IN THE SPECIFICATION

Please amend the second paragraph on page 7 as follows:

To solve the problem of clustering of the magnetic beads the additive is utilized at relatively low concentrations. The final concentration of the additive in step a) of the above described method is in a range of from 2% (w/v) to 7% (w/v). Preferably, the final concentration is in a range of from 3% (w/v) to 6% (w/v), more preferably in a range of from 3% (w/v) to 5% (w/v), and most preferably in a range of from 3% (w/v) to ~~4,8~~4.8% (w/v).

Please amend the first full paragraph on page 17 as follows:

Isolation of gDNA was processed on a BioRobot® 3000 in combination with a BioRobot® RapidPlate™ (both QIAGEN, Hilden, Germany). 50 mg of wheat leaves were frozen in liquid nitrogen and homogenized in a MixerMill MM 300 (Retsch, Haan, Germany). 400 µl of buffer A (~~3,5~~ 3.5 M guanidine isothiocyanate; 25 mM sodium citrate; pH ~~7,0~~ 7.0) were added, thoroughly vortexed, and subsequently centrifuged at 6000 x g for 5 minutes. 200 µl supernatant were transferred into a flat bottom microwell plate. 65 µl of 20 % (w/v) polyethylene glycol 8000 and 20 µl MagAttract® magnetic beads (150 mg / ml; QIAGEN, Hilden, Germany) were added. The solution was mixed and incubated for 5 minutes at room temperature. Magnetic separation was applied and the supernatant was removed. Subsequently, beads were washed by resuspending in 200 µl buffer B (~~1,071~~1.071 M guanidine hydrochloride; ~~107,1~~ 107.1 mM potassium acetate; 50% (v/v) isopropanol; 100 ng/ml RNase A) followed by applying magnetic separation and removing of supernatant. The washing step as described was repeated twice with 100% ethanol. Afterwards, the beads were dried 5 minutes at room temperature. For elution of genomic DNA the beads were resuspended in 100 µl of buffer C (10 mM Tris / HCl; pH ~~8,5~~8.5) and incubated for 5 minutes at room temperature.

Please amend the first paragraph on page 18 as follows:

The isolation was processed on a BioRobot® 3000 in combination with a BioRobot® RapidPlate™ (both QIAGEN, Hilden, Germany). 2 x 2 cm of conifer needles were frozen in liquid nitrogen and homogenized in a MixerMill (Retsch, Haan, Germany). 400 µl of buffer D (~~1,4~~1.4% (w/v) sodium dodecyl sulfate; 50 mM ethylene diamine tetra acetic acid (EDTA); 500 mM sodium chloride; 2% (w/v) polyvinylpyrrolidone; 100mM sodium acetate; pH ~~5,5~~ 5.5) were added, thoroughly vortexed, and subsequently centrifuged at 6000 x g for 5 minutes. The further preparation of the probes was identical to the preparation described in Example 1.

Please amend the third paragraph on page 18 as follows:

30 mg of wheat leaves were homogenized as described above, wherein the added additive was not 65 µl 20% (w/v) polyethylene glycol 8000 but was 50 µl 25% (w/v) of aqueous dextran sulfate. No problems regarding bead handling were observed during the process, i.e. no apparent bead clustering/aggregation was observed.